

Evaluation of a single non-surgical approach in the management of peri-implantitis: glycine powder air-polishing versus ultrasonic device

Lucia Memè¹
Fabrizio Bambini¹
Tommaso Pizzolante²
Francesco Sampalmieri¹
Alessia Bianchi²
Stefano Mummolo²

¹Università Politecnica delle Marche, D.I.S.C.O., Ancona, Italy

²Università degli Studi Dell'Aquila Dipartimento di Medicina Clinica, Scienze della Vita e dell'Ambiente

Corresponding author: Lucia Memè
e-mail: l.meme@staff.univpm.it

Abstract

Background: Precise dimensional impression accuracy is crucial in dental prosthetic. Dental implants are often used to replace lost teeth and present a high level of predictability, patient satisfaction, and long-term success. However, biological complications such as peri-implant mucositis and peri-implantitis have become major challenges to the profession.

Peri-implant mucositis is an inflammation of the soft tissues adjacent to a dental implant diagnosed with bleeding on gentle probing (<0.20 N). If the clinical signs are combined with bone loss, the condition is called peri-implantitis.

The treatment goal of peri-implant disease is to remove or significantly depress the levels of pathogens to allow the healing of the soft and hard tissues. Peri-implant mucositis is a common clinical entity that may develop into peri-implantitis, so early recognition and proper diagnosis of peri-implant disease are highly important in the treatment.

The present study aimed to evaluate the clinical outcome (probing depth PD, bleeding on probing BoP, plaque index PI) following treatment of peri-implantitis with a single non-surgical approach using a glycine powder air-polishing (GPAP) or an ultrasonic device over 2 months.

Thirty implants were enrolled and randomly assigned to test (GPAP) and control (ultrasonic device) groups. Significant differences were found in the mean of the clinical outcome evaluation, but further observations of larger sample size of patients are needed.

Keywords: Peri-implantitis; Dental Implants; Air-Polishing Device; Glycine; Non-Surgical Treatment.

Introduction

The consensus report of the World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Pathological Conditions, which met in 2017 in Chicago, defines peri-implantitis as a biofilm-associated pathological condition involving the peri-implant tissues, characterized by inflammation of the mucosa and progressive loss of bone support (1,2).

Cigarette smoke [3], radiation therapy [4], poor patient compliance [5,6], and susceptibility to periodontal disease are outlined as the main risk factors predisposing to peri-implant disease [8]; recent evidence has also highlighted the role of local factors, such as the width of keratinized mucosa around the implant (9), the excess cement (10) and the prosthetic emergency profile of the reconstruction, which elements exacerbate peri-implant inflammation (12,13).

To date, no reliable evidence suggests the most effective treatment for peri-implantitis (14).

Correct oral hygiene instructions (OHI) are unanimously considered the most necessary

Authors

Lucia Memè - Università Politecnica delle Marche, D.I.S.C.O., Ancona, Italy

Fabrizio Bambini - Università Politecnica delle Marche, D.I.S.C.O., Ancona, Italy

Tommaso Pizzolante - Università degli Studi Dell'Aquila Dipartimento di Medicina Clinica, Scienze della Vita e dell'Ambiente

Francesco Sampalmieri - Università Politecnica delle Marche, D.I.S.C.O., Ancona, Italy

Alessia Bianchi - Università degli Studi Dell'Aquila Dipartimento di Medicina Clinica, Scienze della Vita e dell'Ambiente

Stefano Mummolo - Università degli Studi Dell'Aquila Dipartimento di Medicina Clinica, Scienze della Vita e dell'Ambiente



License

This work is licensed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/).

Authors contributing to Oral and Implantology agree to publish their articles under the [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/), which allows third parties to copy and redistribute the material providing appropriate credit and a link to the license but does not allow to use the material for commercial purposes and to use the material if it has been remixed, transformed or built upon.

How to Cite

L. Memè, F. Bambini, T. Pizzolante, F. Sampalmieri, A. Bianchi, S. Mummolo. Evaluation of a single non-surgical approach in the management of peri-implantitis: glycine powder air-polishing versus ultrasonic device Oral and Implantology Vol. 16 No. 2 (2024), 67-78.

and effective procedure for controlling and reducing oral bacterial biofilm (15). However, the morphological characteristics of implant-supported dental prosthetic rehabilitations can limit the implementation of optimal self-performed plaque control (16).

Whereas the mechanical removal of the biofilm, therefore professional non-surgical therapy in association with adequate home plaque control, proves to be the gold standard in maintaining peri-implant health and treating peri-implant pathologies (17), additional auxiliary therapies can improve clinical outcomes.

The non-surgical treatment of peri-implant disease is generally carried out through correct debridement of the fixture's surface, with the aim of reducing inflammation of the peri-implant tissues (18). Although various mechanical removal methods have been described in the literature, there is no unanimous consensus on defining a preferred decontamination method, which, therefore, remains one of the main topics of discussion (19).

Latest reviews have shown that curettes, ultrasonic instruments, and abrasive powders conveyed with air-polishing devices are the most common instruments used for debridement of the implant surface (20), (21).

Recent in vitro studies, however, have investigated different methods of remediation of implant surfaces affected by peri-implantitis (22), demonstrating that air-polishing devices have greater decontaminating potential than ultrasonic curettes and scalers (23), (24). These air-polishing devices (APD) have been part of everyday clinical practice for many years now as tools that offer a valid therapeutic possibility in maintaining periodontal health (25).

Since the onset of peri-implant mucositis is dependent on biofilm formation, ADPs have the potential also to be used in cases of peri-implantitis (26).

Glycine powder, a non-toxic and water-soluble compound, has been shown not to modify the implant surface profile under scanning electron microscope (SEM) (27).

Several clinical studies have detected a significant improvement in parameters such as probing depth (PD),

bleeding on probing (BoP), and microbiological tests carried out after treatment of implant sites affected by peri-implantitis with glycine powder conveyed via air polishing (GPAP) (28), (29), (30).

However, it is still a topic of discussion that GPAP does not show superior performance when compared to other methodologies, such as manual instrumentation via curettes, ultrasonic scalers, or treatments using YAG (Yttrium Aluminum Garnet) lasers or Erbium lasers (28), (29), (30).

On the other hand, it is also essential to take into consideration the patient's degree of satisfaction with the treatment received in order to determine more precisely whether there is greater comfort and an individual preference in practicing one technique rather than another; several studies highlight how there is a lower degree of discomfort for the patient when undergoing treatment via APD compared to a traditional ultrasound technique (31).

Our study would like to contribute by evaluating the clinical effects on the health of peri-implant tissues in sites affected by peri-implantitis after a single phase of non-surgical treatment using two compared methodologies: glycine powder conveyed via air-polish device (GPAP) and Ultrasonic Instrumentation.

Materials and methods

Study Design

After the application of the eligibility criteria (Table 1), the implants considered suitable n=30 are divided into two groups (Flowchart) and instrumented using the Combi Touch Mectron multifunctional prophylaxis device®:

- TEST GROUP (GPAP) n=15: the implants of the test group are treated with a debridement session via GPAP, explicitly using the Air-Polishing Slim handpiece (Mectron®) with 120° inclination. The PERIO subgingival tip was applied around each side of the implant (mesial, buccal, distal, lingual, or palatal), conveying the glycine powder (Glycine

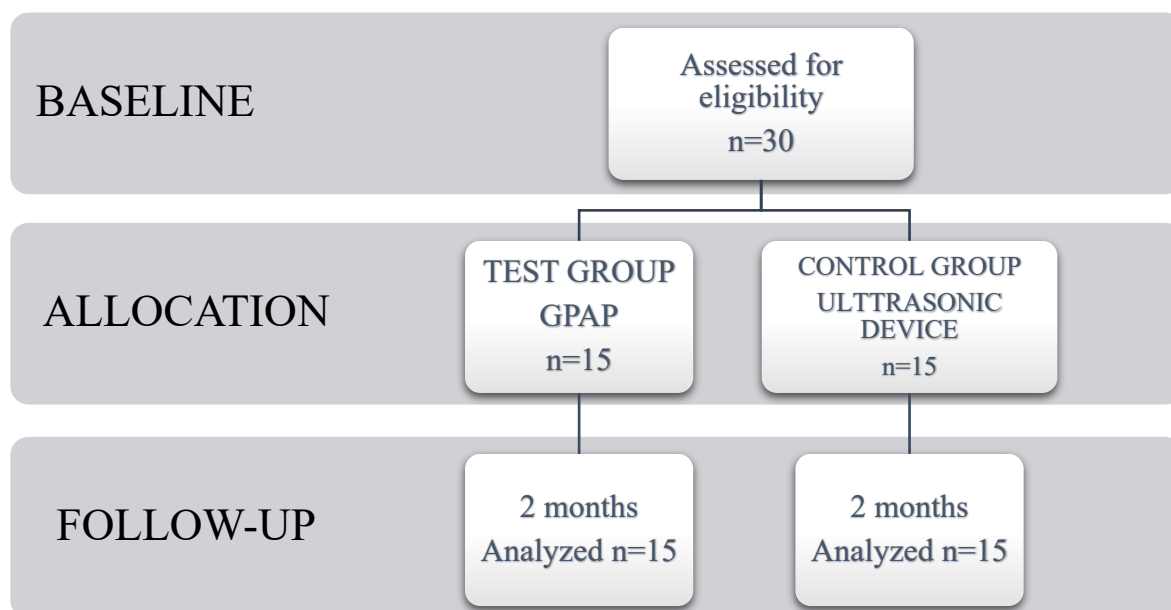
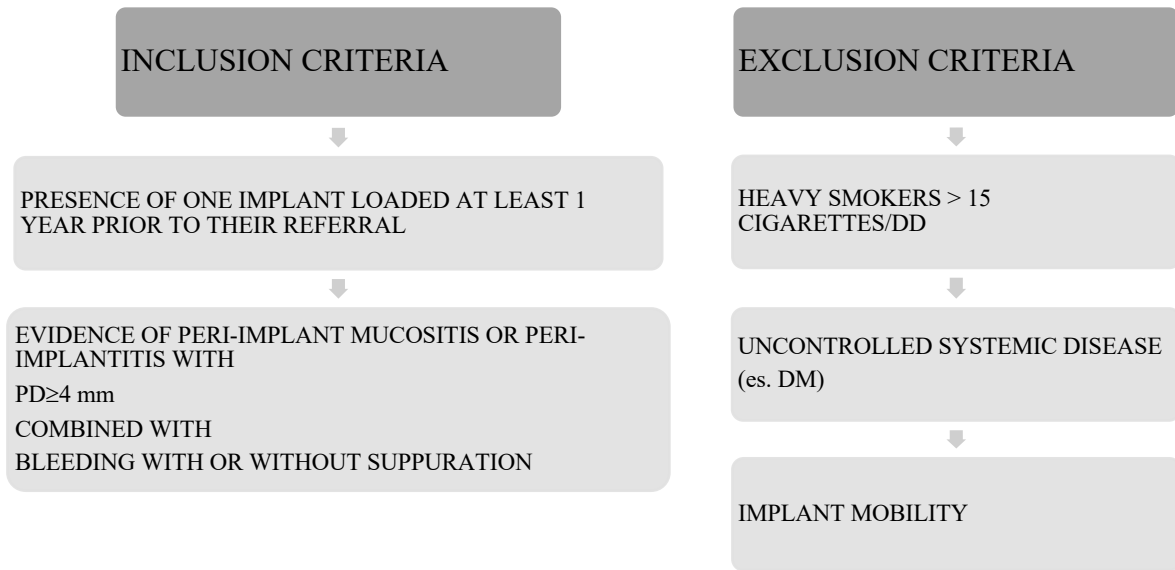


Figure 1. Flowchart of the study protocol

Table 1. Eligibility criteria



Powder, Mectron®) in the sulcus for at least 5 seconds (32), as specified by the manufacturer, or until the clinician deemed the area properly cleaned.

- CONTROL GROUP (ULTRASOUND) n=15: the control group implants are treated using an ultrasound handpiece, Slim Piezoelectric Scaler (Mectron®), with P3 model titanium tip (Perio Universal insert, Mectron®).

Clinical Variables

The clinical parameters of each implant were recorded in the periodontal chart (periodontal chart, Perio-tools®)

at baseline and after two months in re-evaluation (follow-up), accompanied by periapical radiography (Figure 2):

- PD PROBING DEPTH measured as the distance between the gingival margin and the clinical extent of the sulcus/pocket, in six points for each implant
- BOP BLEEDING ON PROBING value 0: no bleeding on probing; value 1: bleeding on positive probing after insertion of the probe.
- PI PLAQUE INDEX value 0: no plaque accumulation; value 1: detection of plaque accumulation using the periodontal probe.

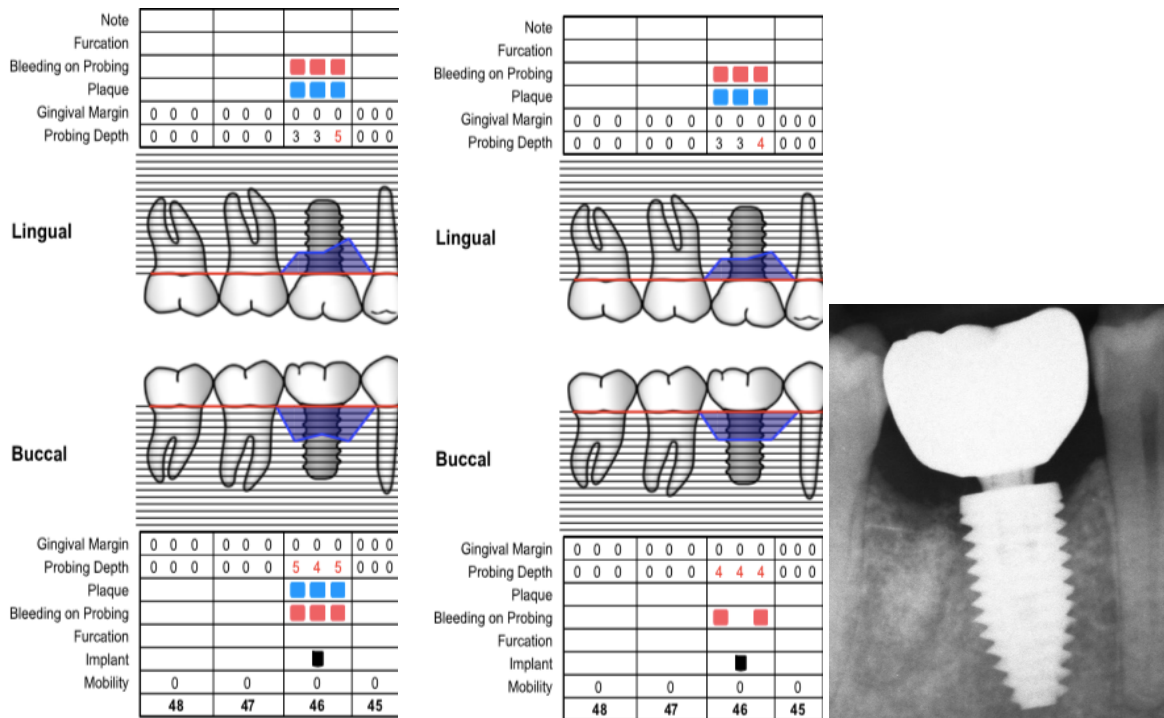


Figure 2. example of data collection in the periodontal chart at baseline and after 2 months (periodontal chart, Perio-tools®) with a digital periapical radiography.

Probing depth, bleeding on probing, and plaque index were measured at 6 implant sites (mesiobuccal, buccal, distobuccal, mesio-lingual or palatal, lingual or palatal, disto-lingual or palatal) using a periodontal probe (UNC 15, HuFriedy®, Chicago, IL, USA) with gentle pressure (approximately 0.20 N).

Randomization

Once the entry criteria had been confirmed, the subjects were entered into the study and assigned an implant number. Assignment to the test group (GPAP) or the control group (ultrasonic device) was made using computerized randomization.

A staff member not involved in the examination or treatment of the implants prepared and placed cards with group identification in numbered envelopes.

The clinician responsible for the treatment broke the

envelope seal to give the therapy according to either group test or control.

Data extraction

n=30 implants divided into 2 groups were analyzed, and the following data were collected

at baseline and 8 weeks after treatment (Tables 2, 3): probing depth (PD), bleeding on probing (BoP), and plaque index (PI). Mean values and standard deviation (mean; SD) for the clinical parameters were calculated for each implant of the two groups.

Following baseline treatment, all patients in both groups were motivated with individual instructions for home oral hygiene (33), with the use of a manual or electric toothbrush, a toothpaste containing sodium fluoride, individualized interdental hygiene devices about the individual width of the interproximal space (34).

Table 2. Baseline and after 2-months clinical variables recorded for implant n=1 in the test group (GPAP)

IMPLANT N° 1	PRE			POST		
	PD	BoP	PI	PD	BoP	PI
SITE						
DV	2	1	1	2	0	0
V	2	1	1	2	0	0
MV	4	1	1	3	1	1
DO	5	1	1	4	1	0
OR	4	1	1	4	1	0
MO	5	1	1	4	1	0
MEAN	3.67	100%	100%	3.17	67%	17%
SD	1.3	0%	0%	0.98	52%	41%

Table 3. Baseline and after 2-months clinical variables recorded for implant n=1 in the control group (ultrasonic device)

IMPLANT N° 1	PRE			POST		
	PD	BoP	PI	PD	BoP	PI
SITE						
DV	3	0	0	3	0	1
V	3	0	0	3	0	0
MV	4	1	1	3	1	0
DO	4	1	1	3	0	0
OR	3	0	0	3	0	0
MO	4	1	1	4	1	1
MEAN	3.5	50%	50%	3.17	33%	33%
SD	0.55	55%	55%	0.41	52%	52%

Statistical analysis

Once the period of collecting clinical parameters at follow-up was completed, the data obtained were entered into a database to obtain a homogeneous and easily findable archive.

The average of the values recorded on the six sites for each implant was calculated and used for all clinical parameters. PD was measured millimeters, while dichotomous variables (BoP and PI) were expressed as percentages (Table 4).

The data were then analyzed, divided into groups based on the relationships to be investigated; the variables were defined using descriptive statistics, mean values, and percentages.

The data has been represented with graphs, histograms, and tables to facilitate understanding.

Results

Sample description

Thirty implants were enrolled in this study. Each implant received treatment according to the randomization. Fifteen implants were treated with the glycine air-polish device (GPAP, test group), and fifteen implants were treated using the ultrasonic device (US, control group). Mean local implant bleeding on probing at baseline was 94,44% in the test group and 91,11% in the control group. Local bleeding on probing varied between the groups during the study period and was reduced between baseline and two months in both groups to 57,78% (test) and 72,22% (control). The mean local plaque index was 93,33% (test) and 92,22% (control) at baseline and was reduced in both groups to 34,44% (test) and 38,89% (control) at the end of the study (Figure 3).

Table 4. summary table of the clinical parameters examined and their variations in the two groups

	TEST GROUP		CONTROL GROUP	
	PRE	POST	PRE	POST
PD	4,044 mm	3,600 mm	3,911 mm	3,733 mm
BoP	94.44%	57.78%	91.11%	72.22%
PI	93.33%	34.44%	92.22%	38.89%

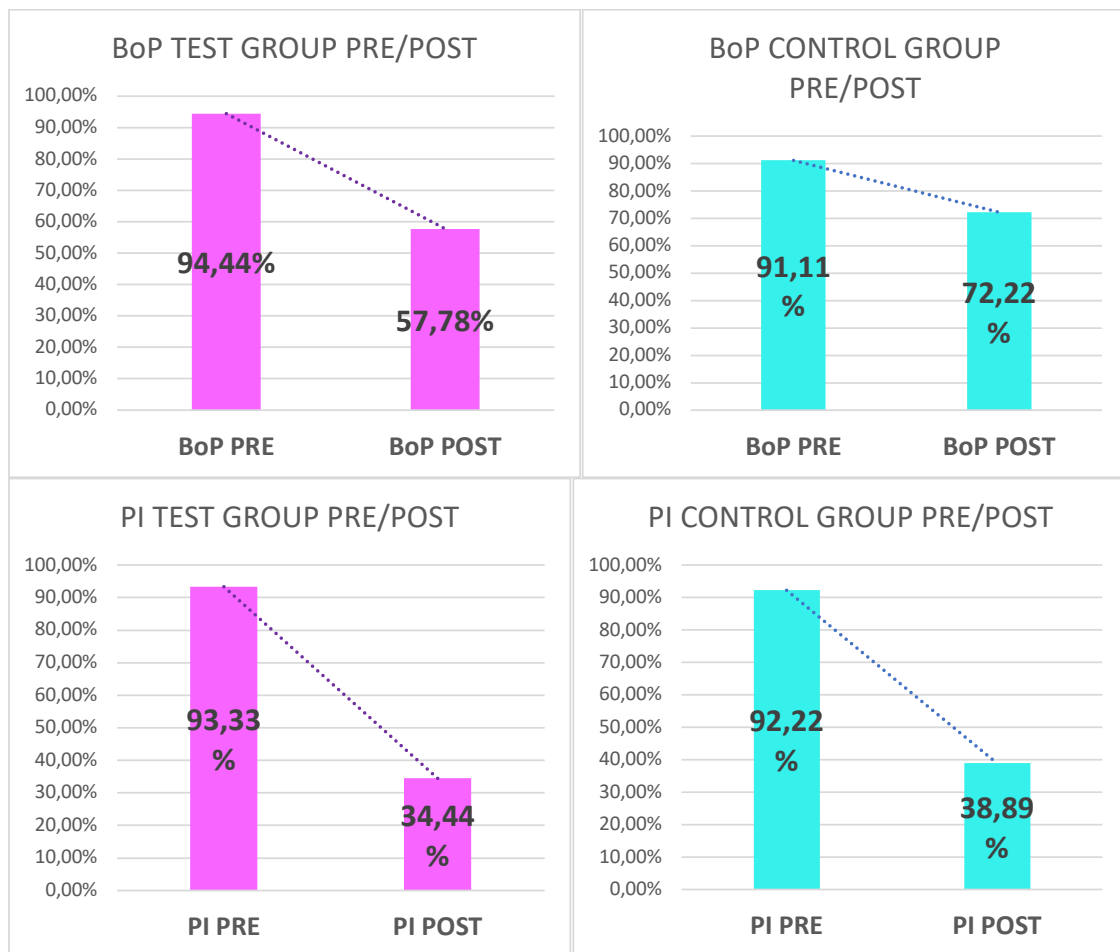


Figure 3. Changes in mean bleeding and plaque score in test and control groups at baseline (pre) and after 2 months (post).

The mean PD was 4,044 mm for test group implants and 3,911 mm for the control group. Mean PD varied between the two groups during the study period and was reduced between baseline and two months in both groups to 3,6 mm in the test group and 3,733 mm in the control group (Figure 4).

The mean PD of each implant was analyzed in both the test and control groups at baseline and after two months (Figs. 5, 6).

Implant pockets were grouped into sites, analyzed for both groups, and divided as follows:

- PD ≤ 2 mm
- 3 mm ≤ PD < 4 mm
- 4 mm ≤ PD < 5 mm
- PD ≥ 5 mm

The percentage of implant pockets in the test group (GPAP) with ≤ 2 mm in probing depth at baseline (pre)

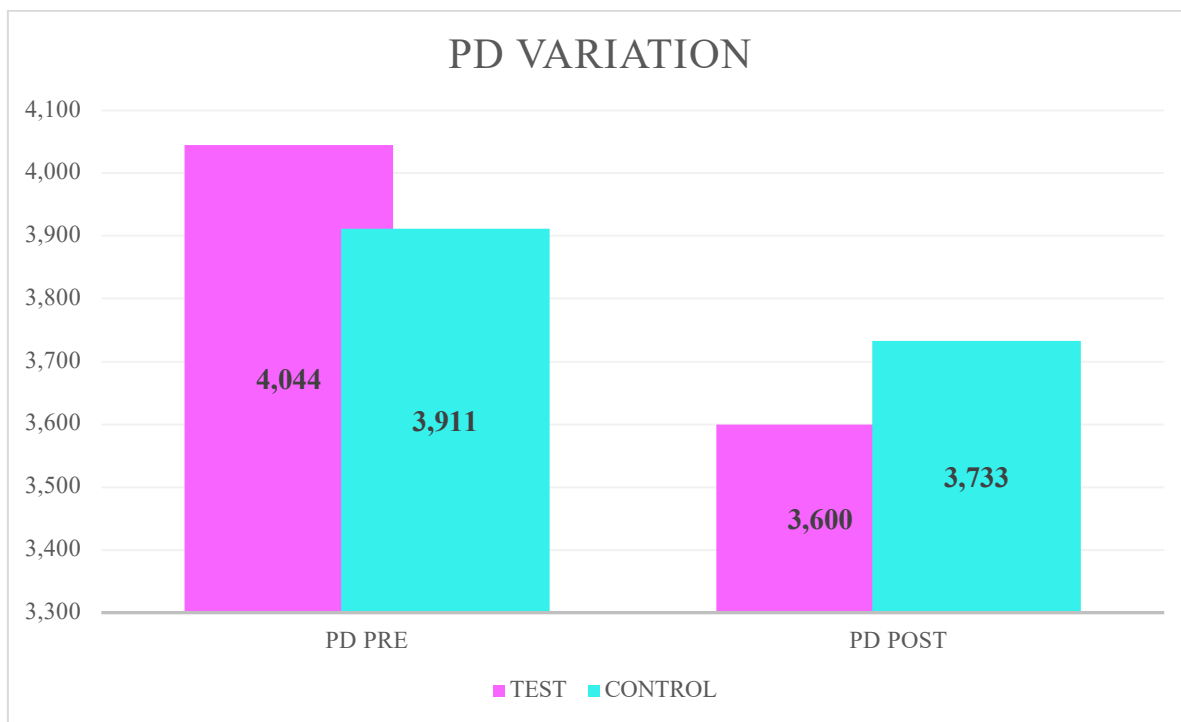


Figure 4. PD changes in mm between the test and control group at baseline and after two months.

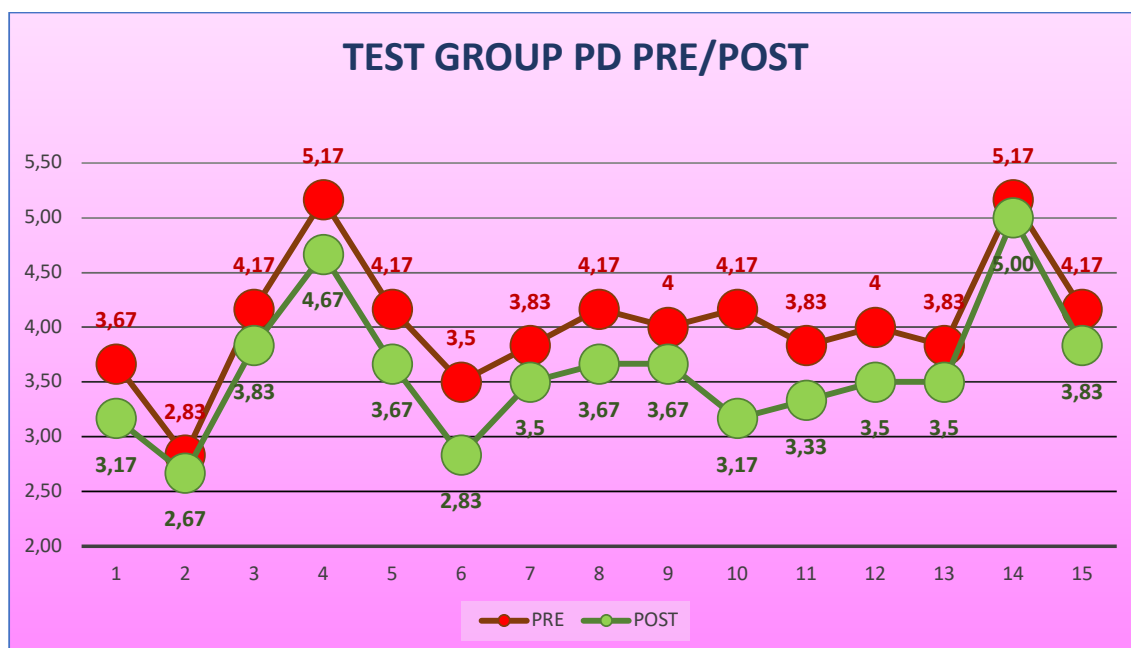


Figure 5. Mean PD variation for each implant at baseline (red) and after two months (green) in the test group (GPAP).

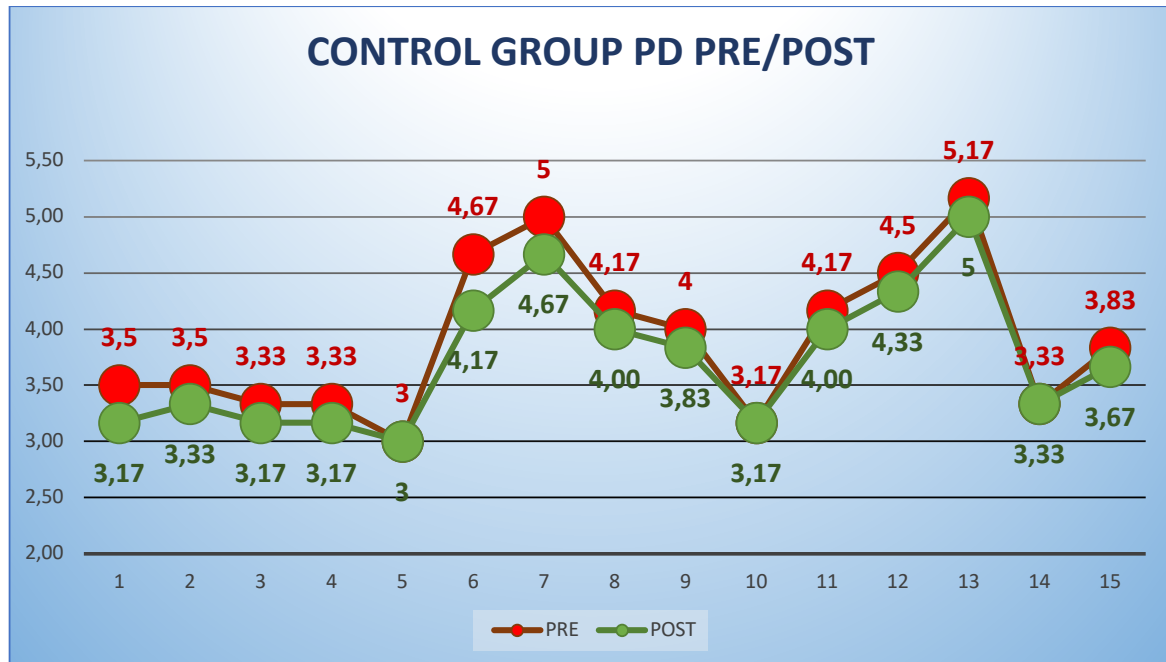


Figure 6. Mean PD variation for each implant at baseline (red) and after two months (green) in the control group (US).

was 5,56% (5 sites), with 3 mm ≤ PD < 4 mm was 21,11% (19 sites), with 4 mm ≤ PD < 5 mm was 41,11% (37 sites) and with PD ≥ 5 mm was 32,22% (29 sites). At the two-month follow-up (post) the number of sites ≤ 2 mm were increased to 12,22% (11 sites), with 3 mm ≤ PD < 4 mm to 30% (27 sites), with 4 mm ≤ PD < 5 mm to 44,44% (40 sites) and the number of sites ≥ 5 mm were reduced to 13,33% (12 sites) respectively (Table 5, Figure 7).

The percentage of implant pockets in the control group (US) with ≤ 2 mm in probing depth at baseline (pre) was 4,44% (4 sites), with 3 mm ≤ PD < 4 mm was 31,11% (28 sites), with 4 mm ≤ PD < 5 mm was 36,67% (33 sites) and with PD ≤ 5 mm was 27,78% (25 sites). At the two-month follow-up (post), the number of sites ≤ 2 mm was reduced to 3,33% (3 sites), with 3 mm ≤ PD < 4 mm were increased to 42,22% (38 sites), with 4 mm ≤ PD < 5 mm reduced to 34,44% (31 sites) and the number of sites ≥ 5 mm were reduced to 20% (18 sites) respectively (Table 6, Figure 8).

No significant difference in the number of sites with 0-4 mm and 4 mm ≤ PD < 5 mm existed between the two groups.

The number of diseased sites (PD ≥ 5 mm) was reduced the most in the test group, from 29 to 12 sites, compared to the control group, from 25 to 18. There was a percentage difference between baseline (pre) and two-month follow-up (post) of 18,89% for the GPAP group and only 7,78% for the ultrasonic group (Figs. 9, 10).

Discussion

The present study reports the results of evaluating a single non-surgical approach to managing peri-implantitis using two different techniques: glycine powder delivered via an air-polishing device (GPAP) and Ultrasonic Instrumentation.

Air-polishing techniques are used in many dentistry fields that require removing bacterial biofilm. They assume considerable importance in the clinical management of periodontitis and peri-implantitis, given that these pathologies are considered a public health problem at a global level. The data available in the literature suggests that the prevalence of peri-implantitis is between 16% and 25% of patients with implants (35).

Table 5. Percentage and number of site variation in PD in the test group (GPAP) between baseline and follow-up at two months.

TEST GROUP	PD PRE		PD POST	
	No	%	No	%
≤ 2mm	5	5.56%	11	12.22%
3mm ≤ PD < 4mm	19	21.11%	27	30.00%
4mm ≤ PD < 5mm	37	41.11%	40	44.44%
≥ 5 mm	29	32.22%	12	13.33%

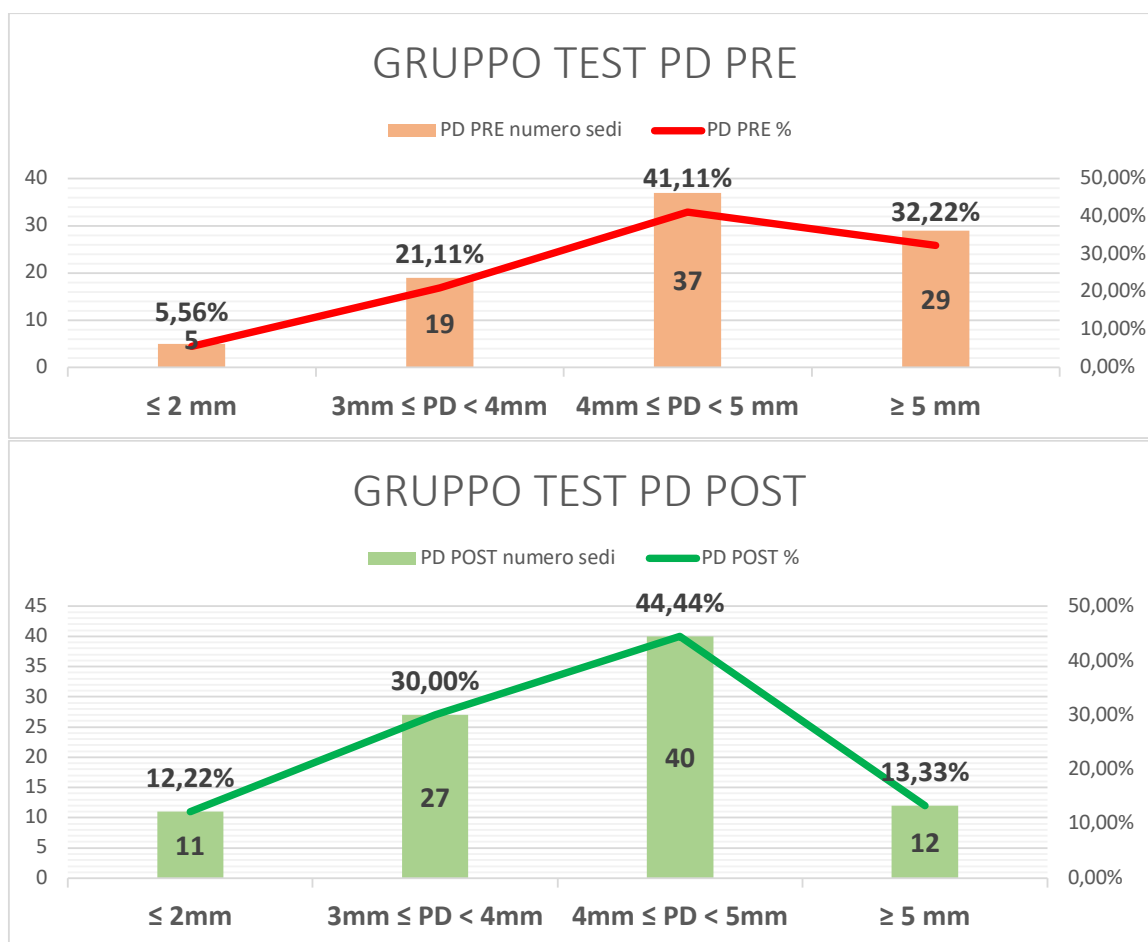


Figure 7. Percentage and number of sites variation in PD in the test group (GPAP) between baseline (red graph) and follow-up at two months (green graph).

Table 6. Percentage and number of site variation in PD in the control group (US) between baseline and follow-up at two months.

CONTROL GROUP	PD PRE		PD POST	
	No	%	No	%
≤ 2mm	4	4.44%	3	3.33%
3mm ≤ PD < 4mm	28	31.11%	38	42.22%
4mm ≤ PD < 5mm	33	36.67%	31	34.44%
≥ 5 mm	25	27.78%	18	20.00%

Many studies have ascertained the cause-effect relationship between the colonization of bacterial plaque and the pathogenesis of peri-implant disease (36). Thus, it is proven that the removal of bacterial biofilm is an essential prerequisite for the management and therapy of peri-implant diseases.

Various techniques have been proposed for treatment, such as mechanical and ultrasonic instrumentation or laser. Air-polishing powder procedures have been widely adopted in the therapy of peri-implantitis, showing no adverse side effects (37-38). On the contrary, several in vitro studies have shown that using substances with high abrasive power (e.g., sodium bicarbonate) can cause alterations in the surface characteristics of the implant

fixtures and on the surface of natural teeth (39).

A less aggressive technique that uses glycine powder expresses excellent effectiveness in Removal of the bacterial biofilm, compared to the methods mentioned above, reduces the biological trauma on the soft tissues, resulting in greater comfort for the patient, and above all, it does not cause alterations to the implant surface (40).

Glycine powder, when compared with other air-polishing powders, does not produce any alteration to the surface characteristics of titanium, as described by many in vitro studies, and this property is not influenced by the distance and angle of the flow of the air polishing (41), (42).

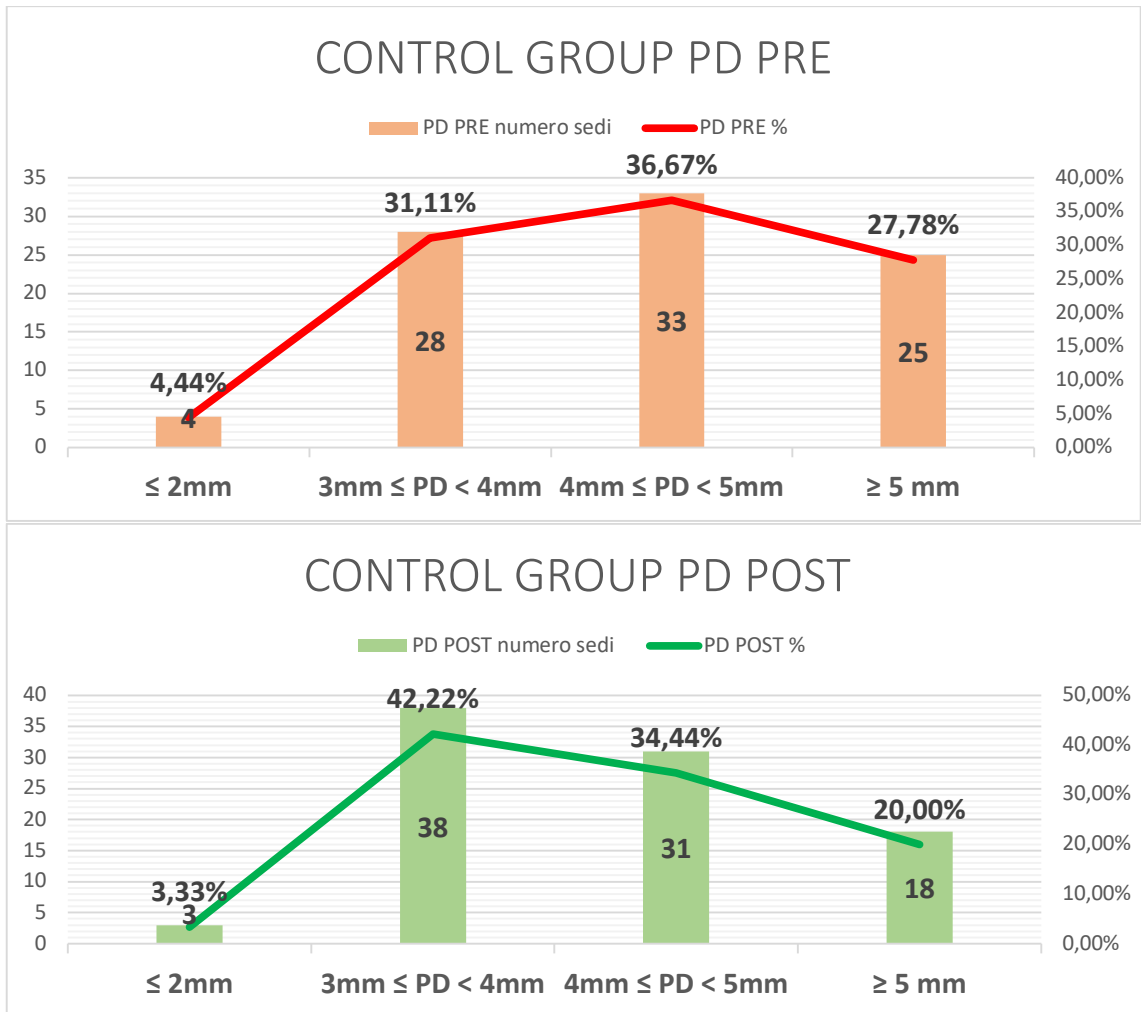


Figure 8. Percentage and number of sites variation in PD in the control group (US) between baseline (red graph) and follow-up at two months (green graph)

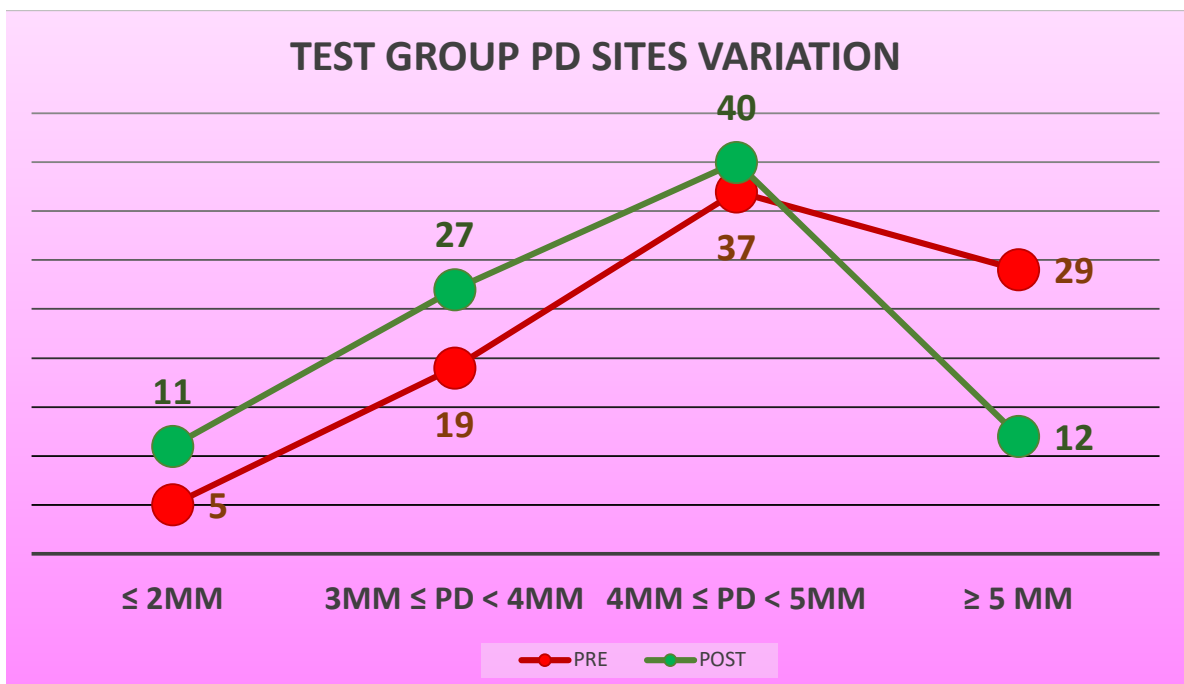


Figure 9. Changes in PD for each analyzed site in test group.

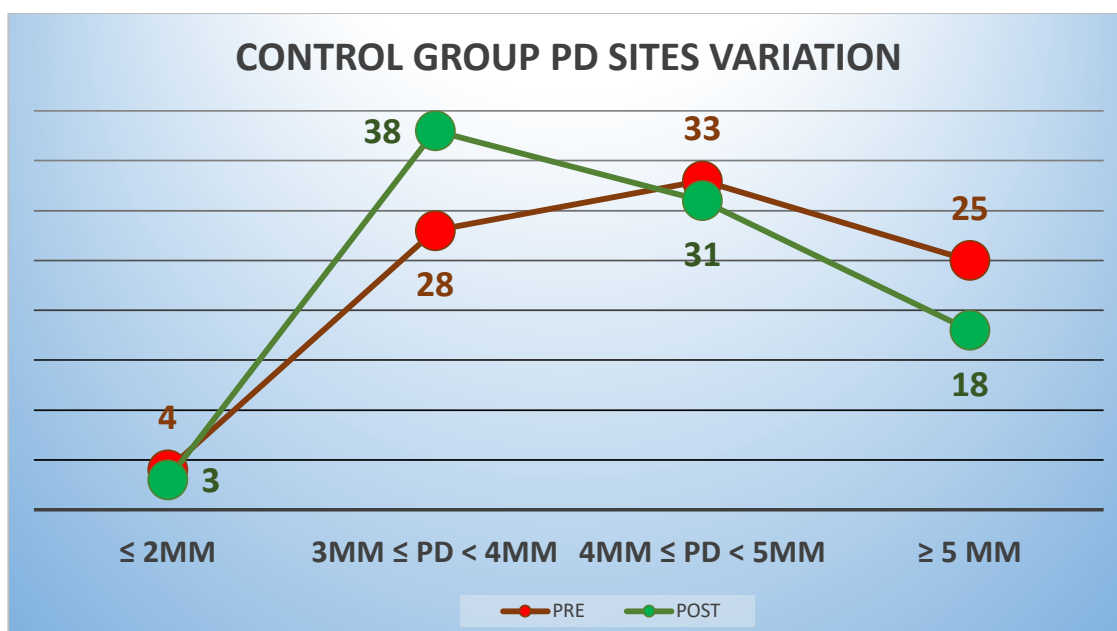


Figure 10. Changes in PD for each analyzed site in control group.

These characteristics, which reduce alterations at the titanium surface level, could lead to less development and formation of biofilm. In this study, a marked difference in PI values is observed between the two groups, despite the baseline measurement being similar (in the test group, the percentage drops from 93.33% to 34.44% with a difference of 58.89%, while in the PI control group it varies from 92.22% to 38.89%, with a difference between before and after of 53.33%). In addition to the absence of superficial alterations, the ability of glycine powder to inhibit the formation of bacterial biofilm should also be further investigated.

The discrepancy noted between the two groups regarding probing depth variation (Fig. 4) could be attributed more to the minimal differences in PD values at baseline. While many studies (43) focus on the treatment of severe peri-implantitis, the present research mostly included patients with initial to moderate lesions while still being in line with the results of many other works that investigated the effectiveness of GPAP for the treatment of peri-implant pathologies (44,45). Ji et al. identified a significant reduction in PD and BoP through this type of treatment, even if they failed to find additional beneficial effects of GPAP compared with traditional ultrasonic instrumentation (46,47). Similarly, Riben-Grundstrom et al. demonstrated in their study a constant improvement in PD and BoP parameters. Still, they suggested that both causal therapy modalities are reliable tools in the non-surgical treatment of peri-implant pathologies (48-51).

Our study has, therefore, demonstrated that professional biofilm removal strategies, in particular the use of glycine powder conveyed via an air-polishing device (GPAP) and associated with professional individualized oral hygiene instructions, reduce the extent of bleeding on probing (BoP) and plaque index (PI) at two months re-evaluation (Figure 11).

To summarize, the present study shows that both clinical procedures can improve inflammatory conditions with a superior benefit in the test group (GPAP) after the two-month follow-up in terms of BoP and PI. Furthermore, the

use of glycine conveyed with an air-polish device (GPAP) leads to a significant improvement in the probing depth over time if compared to the control group (ultrasonic device), and this may be due to a trophic effect of the glycine powder on the peri-implant soft tissues, given by the specific cytoprotective characteristics towards the periodontal tissues (52-55).

In a study by Petersilka et al. (56-57) the author describes the effects on periodontal tissues of air-polishing with glycine powder versus bicarbonate powder and manual instrumentation. Glycine powder has a minimally erosive effect on the gingival epithelium when compared with other methodologies; this research can be considered as a confirmation of the results obtained in our study since a lower traumatic effect on the tissues may have caused the beneficial effects on PD measurements.

Conclusion

GPAP treatment of implant surfaces is, therefore, a promising option in the causal therapy of peri-implantitis and is increasingly used.

The results of the present study demonstrate that it is possible to obtain a significant improvement in sites affected by moderate peri-implantitis using glycine powder, in agreement with many studies carried out in recent years (51), although randomized clinical trials with larger sample sizes are rendered meaningful to further evaluate the long-term effects and peri-implant stability after this non-surgical option treatment in peri-implant disease.

References

1. Berglundh T, Armitage G, Araujo MG, Avila-Ortiz G, Blanco J, Camargo PM, et al. Peri-implant diseases and conditions: Consensus report of workgroup 4 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions. In: Journal of Clinical Perio-dontology. Blackwell Munksgaard; 2018. p. S286-91.

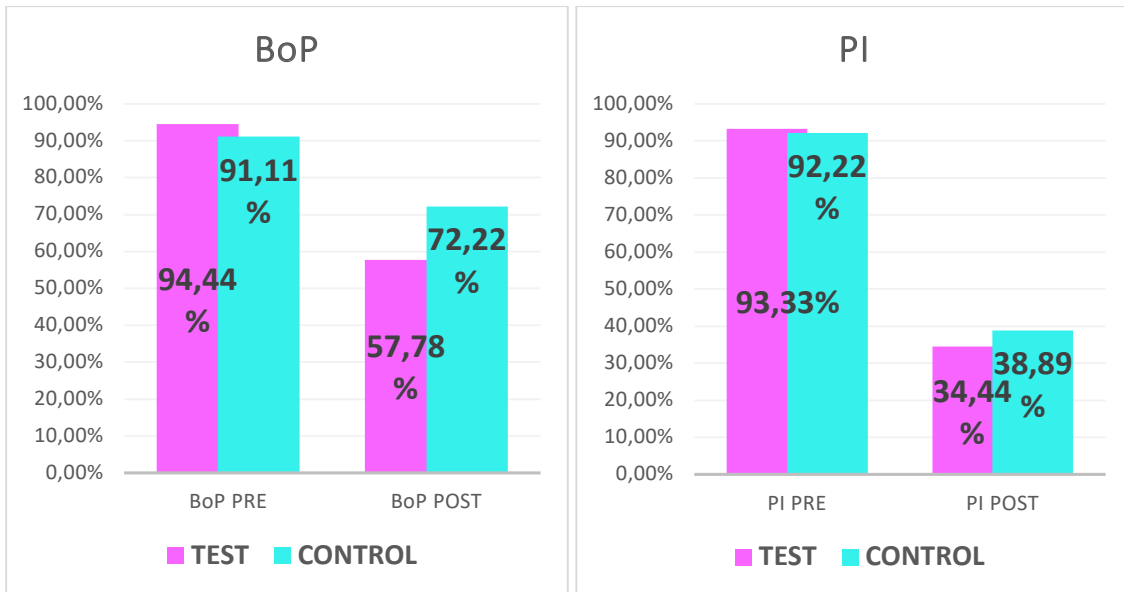


Figure 11. Changes in BoP and PI between test and control group at baseline and after 2-month follow-up.

2. Marchetti, E., Mummolo, S., Mancini, L., ...Marzo, G., Campanella, V. Decontamination in the dental office: a comparative assessment of a new active principle I Decontaminazione nello studio odontoiatrico: Valutazione di un nuovo principio attivo Dental Cadmos, 2021, 89(3), pp. 200–206
3. Martinez-Amargant J, de Tapia B, Pascual A, Takamoli J, Esquinas C, Nart J, et al. Association between smoking and peri-implant diseases: A retrospective study. Clin Oral Implants Res. 2023 Oct 1;34(10):1127–40.
4. Mummolo, S., Severino, M., Campanella, V., ...Quinzi, V., Marchetti, E. Periodontal disease in subjects suffering from coronary heart disease Journal of Biological Regulators and Homeostatic Agents, 2019, 33(3), pp. 73–82
5. Kwon YD, Karbach J, Wagner W, Al-Nawas B. Peri-implant parameters in head and neck reconstruction: Influence of extraoral skin or intraoral mucosa. Clin Oral Implants Res. 2010 Mar;21(3):316–20.
6. Mummolo, S., Nota, A., Marchetti, E., ...Marzo, G., Campanella, V. Histologic and histomorphometric analysis of maxillary sinus augmentation with different biomaterials. A pilot split-mouth human study ORAL and Implantology, 2018, 11(4), pp. 249–256
7. Costa FO, Takenaka-Martinez S, Cota LOM, Ferreira SD, Silva GLM, Costa JE. Peri-implant disease in subjects with and without preventive maintenance: A 5-year follow-up. J Clin Periodontol. 2012 Feb;39(2):173–81.
8. Daubert DM, Weinstein BF, Bordin S, Leroux BG, Flemmig TF. Prevalence and Predictive Factors for Peri-Implant Disease and Implant Failure: A Cross-Sectional Analysis. J Periodontol. 2015 Mar;86(3):337–47.
9. Bambini F, Pellicchia, M., Memè, L., ... Procaccini, M., Lo Muzio, L. Anti-inflammatory cytokines in peri-implant soft tissues: A preliminary study on humans using cDNA microarray technology European Journal of Inflammation, 2007, 5(3), pp. 121–127
10. Boynueğri D, Nemli SK, Kasko YA. Significance of keratinized mucosa around dental implants: A prospective comparative study. Clin Oral Implants Res. 2013 Aug;24(8):928–33.
11. Linkevicius T, Puisys A, Vindasiute E, Linkeviciene L, Apse P. Does residual cement around implant-supported restorations cause peri-implant disease? A retrospective case analysis. Clin Oral Implants Res. 2013 Nov;24(11):1179–84.
12. D'Amario, M., De Angelis, F., Vadini, M., ...Mummolo, S., D'Arcangelo, C. Influence of a repeated preheating procedure on mechanical properties of three resin composites Operative Dentistry, 2015, 40(2), pp. 181–189
13. Bambini, F., Santarelli, A., Putignano, A., ... Emanuelli, M., Lo Muzio, L. Use of supercharged cover screw as static magnetic field generator for bone healing, 1st part: In vitro enhancement of osteoblast-like cell differentiation Journal of Biological Regulators and Homeostatic Agents, 2017, 31(1), pp. 215–220
14. Katafuchi M, Weinstein BF, Leroux BG, Chen YW, Daubert DM. Restoration contour is a risk indicator for peri-implantitis: A cross-sectional radiographic analysis. J Clin Periodontol. 2018 Feb 1;45(2):225–32.
15. Monje A, Insua A, Wang HL. Understanding peri-implantitis as a plaque-associated and site-specific entity: On the local predisposing factors. Vol. 8, Journal of Clinical Medicine. MDPI; 2019.
16. Esposito M, Grusovin MG, Worthington H V. Cochrane Review of Peri-implantitis. Senior Clinical Teaching Fellow in Implant Dentistry [Internet]. Available from: <http://www.cochrane.org>.
17. Van DerWeijden FA, Slot DE. Efficacy of homecare regimens for mechanical plaque removal in managing gingivitis a meta review. Vol. 42, Journal of Clinical Periodontology. Blackwell Munksgaard; 2015. p. S77–91.
18. Pons R, Nart J, Valles C, Salvi GE, Monje A. Self-administered proximal implant-supported hygiene measures and the association to peri-implant conditions. J Periodontol. 2021 Mar 1;92(3):389–99.
19. Barootchi S, Ravidà A, Tavelli L, Wang HL. Nonsurgical treatment for peri-implant mucositis: a systematic review and meta-analysis [Internet]. Vol. 13, Int J Oral Implantol. 2020. Available from: www.crd.york.ac.uk/PROSPERO
20. Muthukuru M, Zainvi A, Esplugues EO, Flemmig TF. Nonsurgical therapy for the management of peri-implantitis: A systematic review. Vol. 23, Clinical Oral Implants Research. 2012. p. 77–83.
21. Máximo MB, De Mendonça AC, Renata Santos V, Figueiredo LC, Feres M, Duarte PM. Short-term clinical and microbiological evaluations of peri-implant diseases before and after mechanical anti-infective therapies. Clin Oral Implants Res. 2009 Jan;20(1):99–108.
22. Schwarz F, Becker K, Renvert S. Efficacy of air polishing for the non-surgical treatment of peri-implant diseases: A systematic review. Vol. 42, Journal of Clinical Periodontology. Blackwell Munksgaard; 2015. p. 951–9.
23. Schwarz F, Schmucker A, Becker J. Efficacy of alternative or adjunctive measures to conventional treatment of peri-implant mucositis and peri-implantitis: a systematic review and meta-analysis. Int J Implant Dent. 2015 Dec;1(1).
24. Giffi R, Pietropaoli D, Mancini L, Tarallo F, Sahrman P, Marchetti E. The efficacy of different implant surface decontamination methods using spectrophotometric

- analysis: an in vitro study. *J Perio-dontal Implant Sci.* 2022;52(6).
25. Ronay V, Merlini A, Attin T, Schmidlin PR, Sahrman P. In vitro cleaning potential of three implant debridement methods. Simulation of the non-surgical approach. *Clin Oral Implants Res.* 2017 Feb 1;28(2):151–5.
 26. Sahrman P, Ronay V, Hofer D, Attin T, Jung RE, Schmidlin PR. In vitro cleaning potential of three different implant debridement methods. *Clin Oral Implants Res.* 2015 Mar 1;26(3):314–9.
 27. Subgingival debridement of periodontal pockets by air polishing in comparison with ultrasonic instrumentation during maintenance therapy. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1600-051X.2011.01751.x>
 28. Sahrman P, Ronay V, Sener B, Jung RE, Attin T, Schmidlin PR. Cleaning potential of glycine air-flow application in an in vitro peri-implantitis model. *Clin Oral Implants Res.* 2013 Jun;24(6):666–70.
 29. Schwarz F, Ferrari D, Popovski K, Hartig B, Becker J. Influence of different air-abrasive powders on cell viability at biologically contaminated titanium dental implants surfaces. *J Biomed Mater Res B Appl Biomater.* 2009 Jan;88(1):83–91.
 30. Máximo MB, De Mendonça AC, Renata Santos V, Figueiredo LC, Feres M, Duarte PM. Short-term clinical and microbiological evaluations of peri-implant diseases before and after mechanical anti-infective therapies. *Clin Oral Implants Res.* 2009 Jan;20(1):99–108.
 31. Renvert S, Lindahl C, Jansåker AMR, Persson RG. Treatment of peri-implantitis using an Er:YAG laser or an air-abrasive device: A randomized clinical trial. *J Clin Periodontol.* 2011;38(1):65–73.
 32. Sahn N, Becker J, Santel T, Schwarz F. Non-surgical treatment of peri-implantitis using an air-abrasive device or mechanical debridement and local application of chlorhexidine: A prospective, randomized, controlled clinical study. *J Clin Periodontol.* 2011 Sep;38(9):872–8.
 33. Bühler J, Amato M, Weiger R, Walter C. A systematic review on the patient perception of periodontal treatment using air polishing devices. *Int J Dent Hyg.* 2016 Feb 1;14(1):4–14.
 34. Riben-Grundstrom C, Norderyd O, André U, Renvert S. Treatment of peri-implant mucositis using a glycine powder air-polishing or ultrasonic device: A randomized clinical trial. *J Clin Periodontol.* 2015 May 1;42(5):462–9.
 35. Bambini F, Santarelli A., Putignano, A., ... Emanuelli, M., Lo Muzio, L. Use of supercharged cover screw as static magnetic field generator for bone healing, 1st part: In vitro enhancement of osteoblast-like cell differentiation *Journal of Biological Regulators and Homeostatic Agents*, 2017, 31(1), pp. 215–220
 36. Van DerWeijden FA, Slot DE. Efficacy of homecare regimens for mechanical plaque removal in managing gingivitis a meta review. Vol. 42, *Journal of Clinical Periodontology*. Blackwell Munksgaard; 2015. p. S77–91.
 37. Gennai S, Nisi M, Perić M, Marhl U, Izzetti R, Tonelli M, et al. Interdental plaque reduction after the use of different devices in patients with periodontitis and interdental recession: A randomized clinical trial. *Int J Dent Hyg.* 2022 May 1;20(2):308–17.
 38. Koldslund OC, Scheie AA, Aass AM. Prevalence of Peri-Implantitis Related to Severity of the Disease With Different Degrees of Bone Loss. *J Periodontol.* 2010 Feb;81(2):231–8.
 39. Mombelli A, Van Oosten MAC, Schiirch E, Lang NP. The microbiota associated with successful healing of osseointegrated titanium implants. Vol. 2, *Oral Microbiol Immunol.* 1987.
 40. Bambini F., Memè L., Pellecchia, M., Sabatucci, A., Selvaggio, R. Comparative analysis of deformation of two implant/abutment connection systems during implant insertion. An in vitro study *Minerva stomatologica*, 2005, 54(3), pp. 129–138
 41. Campanella, V., Mummolo, S., Grazzini, F., Barlattani, A., Di Girolamo, M. The effectiveness of endodontic sealers and endodontic medicaments on the elimination of *Enterococcus faecalis*: An in vitro study. *Journal of Biological Regulators and Homeostatic Agents*, 2019, 33(3), pp. 97–102
 42. Bambini F., Memè L., Procaccini M., Lo Muzio L. Raloxifene covalently bonded to titanium implants by interfacing with (3-aminopropyl)-triethoxysilane affects osteoblast-like cell gene expression *International Journal of Immunopathology and Pharmacology*, 2006, 19(4), pp 905–914
 43. Duarte PM, deMendonça AC, Máximo MBB, Santos VR, Bastos MF, Nociti FH. Effect of Anti-Infective Mechanical Therapy on Clinical Parameters and Cytokine Levels in Human Peri-Implant Diseases. *J Periodontol.* 2009 Feb;80(2):234–43.
 44. Yen Nee W, Raja Awang RA, Hassan A. Effects on the Titanium Implant Surface by Different Hygiene Instrumentations: A Narrative Review. *Cureus.* 2022 Oct 31;
 45. Flemmig TF, Arushanov D, Daubert D, Rothen M, Mueller G, Leroux BG. Randomized Controlled Trial Assessing Efficacy and Safety of Glycine Powder Air Polishing in Moderate-to-Deep Periodontal Pockets. *J Periodontol.* 2012 Apr;83(4):444–52.
 46. Cochis A, Fini M, Carrassi A, Migliario M, Visai L, Rimondini L. Effect of air polishing with glycine powder on titanium abutment surfaces. *Clin Oral Implants Res.* 2013 Aug;24(8):904–9.
 47. Schwarz F, Ferrari D, Popovski K, Hartig B, Becker J. Influence of different air-abrasive powders on cell viability at biologically contaminated titanium dental implants surfaces. *J Biomed Mater Res B Appl Biomater.* 2009 Jan;88(1):83–91.
 48. How do implant surface characteristics influence peri-implant disease? Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1600-051X.2010.01661.x>
 49. Verket A, Koldslund OC, Bunæs D, Lie SA, Romandini M. Non-surgical therapy of peri-implant mucositis—Mechanical/physical approaches: A systematic review. Vol. 50, *Journal of Clinical Periodontology*. John Wiley and Sons Inc; 2023. p. 135–45.
 50. Arcuri, C., Petro, E., Sollecchia, G., Mummolo, S., Marzo, G. Laser in periodontal pockets: In vivo and in vitro study. *Journal of Biological Regulators and Homeostatic Agents*, 2020, 34(3), pp. 139–146
 51. Bambini, F., Giannetti, L., Memè, L., Pellecchia, M., Selvaggio, R. Comparative analysis of direct and indirect stamp impression techniques: An in vitro study *Minerva Stomatologica*, 2005, 54(6), pp. 395–402
 52. Bernardi, S., Mummolo, S., Zeka, K., ...Continenza, M.A., Marzo, G. Use and Evaluation of a Cooling Aid in Laser-Assisted Dental Surgery: An Innovative Study. *Photomedicine and Laser Surgery*, 2016, 34(6), pp. 258–262
 53. Ji YJ, Tang ZH, Wang R, Cao J, Cao CF, Jin LJ. Effect of glycine powder air-polishing as an adjunct in the treatment of peri-implant mucositis: A pilot clinical trial. *Clin Oral Implants Res.* 2014;25(6):683–9.
 54. Riben-Grundstrom C, Norderyd O, André U, Renvert S. Treatment of peri-implant mucositis using a glycine powder air-polishing or ultrasonic device: A randomized clinical trial. *J Clin Periodontol.* 2015 May 1;42(5):462–9.
 55. Breivik T, Gundersen Y, Fonnum F, Vaagenes P, Opstad PK. Chronic glycine treatment inhibits ligature-induced periodontal disease in Wistar rats. *J Periodontol Res.* 2005 Feb;40(1):43–7.
 56. Petersilka G, Faggion CM, Stratmann U, Gerss J, Ehmke B, Haeberlein I, et al. Effect of glycine powder air-polishing on the gingiva. *J Clin Periodontol.* 2008 Apr;35(4):324–32.
 57. Máximo MB, De Mendonça AC, Renata Santos V, Figueiredo LC, Feres M, Duarte PM. Short-term clinical and microbiological evaluations of peri-implant diseases before and after mechanical anti-infective therapies. *Clin Oral Implants Res.* 2009 Jan;20(1):99–108.